Effect of the novel wound healing agent, OPAL A on leukotriene B₄ production in human neutrophils and 5-lipoxygenase activity

Russell FD, Windegger T, Hamilton KD & Cheetham NWH

Abstract

OPAL A is a papaya pulp that is heated and alkalised with bicarbonate (the OPAL process) and is undergoing clinical trials for treatment of chronic wounds. The aim of this study was to investigate possible inhibitory effects of OPAL A and a non-alkalised papaya filtrate on the 5-lipoxygenase signalling pathway. Human isolated neutrophils were incubated with or without OPAL A, non-alkalised papaya or sodium bicarbonate and then exposed to the calcium ionophore, ionomycin to stimulate leukotriene B_4 (LTB₄) production. The production of LTB₄ was inhibited in a dose-dependent manner by all three treatments. The effect of these treatments on 5-lipoxygenase activity, the enzyme involved in the production of precursors of LTB₄ was investigated using a cell-free assay. 5-Lipoxygenase activity was inhibited by OPAL A and non-alkalised papaya, but not bicarbonate. Column chromatography was used to show that the active components within OPAL A were non-proteinaceous polar compounds. The inhibitory effects of OPAL A and a non-alkalised papaya filtrate on 5-lipoxygenase activity and LTB₄ production suggest a possible anti-inflammatory mode of action.

Keywords: papaya, OPAL A, wound healing, 5-lipoxygenase activity, leukotriene B₄.

Introduction

Inflammation is a response to cellular injury and results in the killing of microbial pathogens and tissue destruction. Resolution involves a switch from a pro-inflammatory phase

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that produces mediators such as leukotriene (LT) B_4 to a proresolving phase where resolvins and protectins are produced ¹⁻³. Following activation of neutrophils, $[Ca^{2+}]i$ is elevated, initiating the translocation of 5-lipoxygenase to the perinuclear envelope where it interacts with a 5-lipoxygenase activating protein and arachidonic acid to produce LTA_4 , the precursor of $LTB_4^{-4.5}$. LTB_4 activates BLT_1 and BLT_2 receptors to elicit recruitment and penetrative transmigration of neutrophils from the postcapillary venule, and prolongation of neutrophil survival ⁶⁻⁹. On the basis of such multifaceted involvement in inflammation, BLT_1 and BLT_2 receptors and the synthetic 5-lipoxygenase signalling pathway have become potential therapeutic targets in the management of conditions in which inflammation is implicated ¹⁰⁻¹².

OPAL A is a filtrate that has been manufactured following the homogenisation, heat treatment, alkalisation and filtration of the pulp of the ripened fruit of *Carica Papaya* (the OPAL process), and is currently being examined in a clinical trial conducted by Phoenix Eagle Company in patients who have non-healing wounds (ClinicalTrials.gov Identifier NCT00933348). The mechanisms by which OPAL A contributes to wound healing are not known. We have previously reported a nitric oxide-dependent vasorelaxant effect of OPAL A and a non-alkalised papaya filtrate which raised the possibility that OPAL A might improve perfusion within the wound region¹³. The aim of this study is to examine possible inhibitory effects of OPAL

A and a non-alkalised papaya filtrate on LTB₄ production and 5-lipoxygenase activity.

Materials and methods

Isolation of human neutrophils

Whole blood was obtained from the antecubital vein of five healthy men (22-55 years) and placed in EDTA tubes. Neutrophils were isolated by differential centrifugation using polymorphprep according to manufacturer's instructions (Axis-Shield, Oslo, Norway). Briefly, whole blood was layered onto polymorphprep in a 1:1 ratio and centrifuged at 500xg for 30 minutes at 22°C in an Eppendorf Centrifuge 5702 with a swing rotor. The fraction containing neutrophils was collected and diluted with 8 ml M199 culture media containing 20% fetal calf serum, 2 mM Glutamax-1, 2.5 μg/ml fungizone and 50 μg/ml penicillin/streptomycin (M199 media), and spun at 450xg for 30 minutes at 22°C. The cell pellet was resuspended in 450 µl Hanks' Balanced Salt Solution (HBSS), with 10 µl of sample smeared onto a microscope slide and stained using Diff Quik differential dye to confirm successful isolation of neutrophils (Figure 1A). This study was conducted in 2010-2011 and conforms to the Statement on Human Experimentation and was carried out with approval from the Human Research Ethics Committee of the University of the Sunshine Coast (A/10/243).

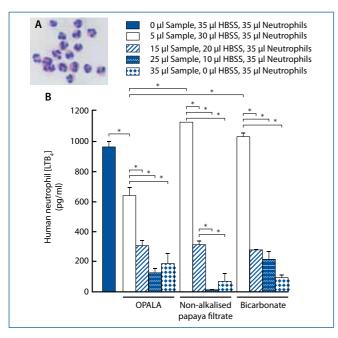


Figure 1. Effect of OPAL A, non-alkalised papaya filtrate and sodium bicarbonate on ionomycin-stimulated production of LTB_4 in isolated human neutrophils. Successful isolation of neutrophils was achieved by differential centrifugation, as indicated by staining using Diff Quik differential dye (A). OPAL A was more potent than non-alkalised papaya and sodium bicarbonate for inhibition of LTB_4 production (B). Values are mean \pm SEM, n=4-5, \star , P<0.05.

Effect of OPAL A and non-alkalised papaya filtrate on LTB₄ concentration in ionomycin-stimulated human neutrophils

Aliquots of resuspended cells (35 μ l) were combined with OPAL A, non-alkalised OPAL A or 10% bicarbonate (5-35 μ l), with solutions prepared to a final volume of 70 μ l with HBSS. Samples were incubated for 20 minutes at 37°C in a 5% CO $_2$ incubator, and then incubated for a further 5 minutes at 37°C with 2 μ M ionomycin before centrifugation at 40xg for 6 minutes at 22°C in a MiniSpin Plus centrifuge (Eppendorf). LTB $_4$ concentration was determined spectrophotometrically at wavelength 405 nm using 50 μ l aliquots of the supernatant in a LTB $_4$ enzyme immunoassay according to manufacturer's instructions (Cayman Chemical Company, Ann Arbor, USA). Background absorbance was measured in the absence of OPAL A, non-alkalised papaya filtrate or bicarbonate and this was subtracted from all readings.

Effect of OPAL A and non-alkalised papaya filtrate on 5-lipoxygenase activity using a cell-free assay

The effect of OPAL A and non-alkalised papaya filtrate on lipoxygenase activity was determined using a modified method of Anthon and Barrett (2001)¹⁴. Lipoxygenase enzyme (3 $\mu g/ml$) was incubated with 10 mM 3-(dimethylamino) benzoic acid, 0.05 M Na₂HPO₄.12H₂O (pH 6.0), 0.5 mM linoleic acid and either OPAL A, non-alkalised papaya filtrate or bicarbonate (100 $\mu l/0.5$ ml solution) (Solution A) for 5 minutes at 22°C. Solution B (0.5 ml), containing 10 mM 3-methyl-2-benzothiazolinone and 0.1 mg/ml haemoglobin was added to 0.5 ml of Solution A, and incubated for a further 5 minutes at 22°C. An aliquot of the mixture was measured using a spectrophotometer at 598 nm.

Column fractionation of OPAL A

To further characterise the inhibitory activity of OPAL A, a 1.5 ml aliquot of OPAL A was passed through a 0.45 µm syringe filter and the filtrate (0.6 ml) was loaded onto an OASIS column that was pre-activated by application of methanol then distilled water. The column was eluted using a mixture of methanol and water (1:1), and the eluant was collected and analysed using the 5-lipoxygenase activity assay described above. In a second series of experiments, 7.0 ml of OPAL A was mixed with 5 g of silica gel. The sample was frozen at -80°C, freeze-dried, then added to the top of a column containing 5 g silica gel that was moistened with ethyl acetate. The column was eluted with successive rinses with 40 ml of ethyl acetate, acetone, methanol and water. The ethyl acetate, acetone and methanol fractions were recovered by evaporation of solvent at 22°C under a stream of nitrogen. The aqueous fraction was freeze-dried. Samples were reconstituted in 0.5 ml phosphate buffer solution (pH 9) and analysed using the lipoxygenase activity assay as described above.

Statistics

Data were compared using one way Analysis of Variance with Tukey's Honestly Significant Difference (HSD) test using IBM SPSS Statistics Version 19. Data are expressed as mean±SEM.

Results

Effect of OPAL A and non-alkalised papaya filtrate on leukotriene B₄ production in ionomycin-activated neutrophils

OPALA, non-alkalised papaya filtrate and sodium bicarbonate inhibited the ionomycin-stimulated production of LTB $_4$ in isolated human neutrophils. The lowest amount of OPALA used in the assay (5 μl / 35 μl of cells) caused a significant reduction in LTB $_4$ production compared to an untreated control, and this was maximal for 15 μl of OPALA/35 μl of cells (Figure 1B). The same amount of non-alkalised papaya filtrate or 10% sodium bicarbonate did not inhibit LTB $_4$ production. Significant inhibition of LTB $_4$ production was observed using 15 μl of non-alkalised papaya or 10% sodium bicarbonate / 35 μl of cells, and this was maximal with 25 μl of non-alkalised papaya or 15 μl of 10% sodium bicarbonate /35 μl of cells.

Effect of OPAL A and non-alkalised papaya filtrate on 5-lipoxygenase activity

OPAL A and the non-alkalised papaya filtrate produced concentration-dependent inhibition of 5-lipoxygenase activity. The equivalent amount of sodium bicarbonate to that contained within each OPAL A solution had no effect on 5-lipoxygenase activity (Figure 2A). Further characterisation of the 5-lipoxygenase activity was achieved by loading OPAL A onto an OASIS column and eluting it with a 1:1 mixture of methanol and water. The dried and reconstituted eluant retained the capacity for inhibition of 5-lipoxygenase activity that was observed in the non-fractionated OPAL A sample (Figure 2B). A further experiment eluted OPAL A from a silica gel column using successive rinses with ethyl acetate, acetone, methanol, and water. The reconstituted eluants obtained using ethyl acetate and acetone had no inhibitory effect on 5-lipoxygenase activity whereas inhibitory activity was obtained with the reconstituted eluants obtained using methanol and water (Figure 2C).

Discussion

Papaya latex harvested from unripe papaya fruit stimulates a pro-inflammatory response when injected into rat paw¹⁵. In this study we examined the possible anti-inflammatory effects of non-alkalised papaya and papaya that was prepared by homogenisation, heat treatment, alkalisation and filtration of the pulp of ripened papaya (the OPAL process). To our

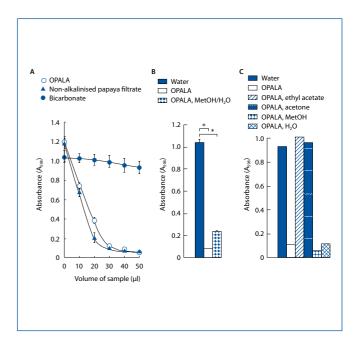


Figure 2. Effect of OPAL A, non-alkalised papaya filtrate and sodium bicarbonate on 5-lipoxygenase activity in a cell-free assay (n=3). OPAL A and non-alkalised papaya inhibited 5-lipoxygenase activity in a dose-dependent manner, whereas sodium bicarbonate was without effect (A). OPAL A was loaded onto an OASIS column and eluted with methanol/water (1:1) (n=3; duplicate samples). The dried and reconstituted (phosphate buffer solution) eluant inhibited 5-lipoxygenase activity (B). OPAL A was mixed with silica gel and loaded onto silica gel columns and eluted with sequential washes with ethyl acetate, acetone, methanol, and water (n=1, duplicate samples). The dried and reconstituted (phosphate buffer solution) eluants were either without effect (ethyl acetate and acetone eluants) or inhibited 5-lipoxygenase activity (methanol and water eluants) (C). Values are mean±SEM (A, B), *, P<0.05.

knowledge, this is the first report identifying the inhibitory action of papaya-based filtrates on the 5-lipoxygenase – ${\rm LTB}_4$ signalling pathway.

OPAL A, non-alkalised papaya and bicarbonate alone inhibited production of LTB₄ by human neutrophils that were exposed to ionomycin. Significant inhibition of LTB₄ production occurred at a lower concentration of OPAL A than for either the non-alkalised papaya or the equivalent amount of bicarbonate. Since OPAL A contains both papaya and sodium bicarbonate, this finding suggests a possible additive inhibitory action on LTB₄ production. LTB₄ is an integral eicosanoid in the inflammatory response, with roles in recruitment and activation of leukocytes, and prevention of leukocyte apoptosis⁷. Thus, inhibition of LTB₄ production by OPAL A suggests a possible mode of action for this filtrate in the treatment of inflammatory conditions.

The endogenous pathway for the synthesis of LTB₄ from arachidonic acid is well described⁴. 5-Lipoxygenase activity is crucial to the production of LTB₄, first catalysing the oxidation of arachidonic acid to generate 5-hydroperoxyeicosatetraenoic acid, then the subsequent production of LTA₄. LTA₄ is in turn converted to LTB₄ in the presence of LTA₄ hydrolase⁴. In the present study, we showed that OPAL A and non-alkalised papaya inhibited 5-lipoxygenase activity with similar potency. Interestingly, sodium bicarbonate was without effect, contrasting with the ability of bicarbonate ions to inhibit LTB₄ production in the activated neutrophils. The findings indicate that bicarbonate, or the elevation of pH resulting from the addition of bicarbonate, may have a direct suppressive effect on the neutrophils.

OPAL A was eluted from an OASIS column or silica gel column as a preliminary analysis of the chemical properties of the filtrate for inhibition of 5-lipoxygenase activity. The eluant obtained from the methanol/water rinse of the OASIS column retained inhibitory activity that was detected in the nonfractionated OPAL A. The polar solvent mixture (methanol/ water) was used to selectively elute polar molecules. When OPAL A was eluted on a silica gel column using nonpolar solvents (ethyl acetate or acetone), inhibitory activity was lost. However, elution with polar solvents (methanol or water) retained activity that was detected in the nonfractionated OPAL A. Proteins efficiently adsorb to silica gel¹⁶, so we conclude that the active components within OPAL A are non-proteinaceous, polar molecules. Bioactivity-guided fractionation experiments could be carried out in the future to elucidate the identity of the active compound(s) present in OPAL A. Phenolic compounds such as quercetin and caffeic acid are candidates as they are 1) polar 2) are expressed in Carica Papaya¹⁷, and inhibit 5-lipoxygenase activity (IC₅₀ for quercetin, 0.6 μM^{18} ; IC₅₀ for caffeic acid, 25 μM^{19}) and LTB₄ production in leukocytes (IC $_{50}$ for quercetin, 2 μM^{20} ; IC $_{50}$ for caffeic acid, 200 µM²¹).

In conclusion, this study has identified an inhibitory effect of OPAL A and non-alkalised papaya filtrate on LTB₄ production and 5-lipoxygenase activity. The findings provide a potential mechanism by which OPAL A might contribute to the treatment of conditions in which inflammation is implicated, such as the treatment of wounds.

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Conflict of interest

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References

- Ariel A & Serhan CN. Resolvins and protectins in the termination program of acute inflammation. Trends Immunol 2007; 28:176–83.
- Kalsotra A, Du L, Wang Y et al. Inflammation resolved by retinoid X receptor-mediated inactivation of leukotriene signaling pathways. FASEB J 2008; 22:538–47.
- Serhan CN, Chiang N & Van Dyke TE. Resolving inflammation: dual antiinflammatory and pro-resolution lipid mediators. Nat Rev Immunol 2008; 8:349–61
- Murphy RC & Gijón MA. Biosynthesis and metabolism of leukotrienes. Biochem J 2007; 405:379–95.
- Evans JF, Ferguson AD, Mosley RT & Hutchinson JH. What's all the FLAP about?: 5-lipoxygenase-activating protein inhibitors for inflammatory disease. Trends Pharmacol Sci 2008; 29:72–8.
- 6. Camp RDR, Coutts AA, Greaves MW, Kay AB & Walport MJ. Responses of human skin to intradermal injection of leukotrienes $\rm C_4$, $\rm D_4$ and $\rm B_4$. Brit J Pharmacol 1983; 80:497–502.
- 7. Tager AM & Luster AD. BLT1 and BLT2: the leukotriene $\rm B_4$ receptors. Prostaglandins Leukotrienes Essent Fatty Acids 2003; 69:123–34.
- Cera MR, Fabbri M, Molendini C et al. JAM-A promotes neutrophil chemotaxis by controlling integrin internalization and recycling. J Cell Sci 2009: 122:268–77.
- Monteiro AP, Pinheiro CS, Luna-Gomes T et al. Leukotriene B4 mediates neutrophil migration induced by heme. J Immunol 2011; 186:6562–7.
- Manigrasso MB & O'Connor JP. Accelerated fracture healing in mice lacking the 5-lipoxygenase gene. Acta Orthopaedica 2010; 81:748–55.
- Saiwai H, Ohkawa Y, Yamada H et al. The LTB4-BLT1 axis mediates neutrophil infiltration and secondary injury in experimental spinal cord injury. Am J Pathol 2010; 176:2352–66.
- Nancey S, Boschetti G, Hacini F et al. Blockade of LTB₄/BLT₁ pathway improves CD8* T-cell-mediated colitis. Inflamm Bowel Dis 2011; 17:279–88.
- 13. Mitchell G, Windegger T & Russell FD. Clinical observations and physiological data supporting a vascular response as a mechanism of the novel wound-healing agent, OPAL A. The 2nd Meeting of the Australasian Wound Tissue Repair Society; 2010 Mar 22-24; Perth (WA), 2010:34.
- Anthon GE & Barrett DM. Colorimetric method for the determination of lipoxygenase activity. J Agric Food Chem 2001; 49:32–7.
- Gupta OP, Sharma N & Chand D. A sensitive and relevant model for evaluating anti-inflammatory activity – papaya latex-induced rat paw inflammation. J Pharmacol Toxicol Meth 1992; 28:15–9.
- Alloue WAM, Destain J, El Medjoub T, Ghalfi H, Kabran P & Thonart P. Comparison of *Yarrowia lipolytica* lipase immobilization yield of entrapment, adsorption, and covalent bond techniques. Appl Biochem Biotechnol 2008; 150:51–63.
- 17. Rivera-Pastrana DM, Yahia EM & González-Aguilar. Phenolic and carotenoid profiles of papaya fruit (*Carica papaya* L.) and their contents under low temperature storage. J Sci Food Agric 2010; 90:2358-65.
- 18. Schewe T, Kühn H & Sies H. Flavonoids of cocoa inhibit recombinant human 5-lipoxygenase. J Nutr 2002; 132:1825–9.
- Doiron J, Boudreau LH, Picot N, Villebonet B, Surette ME & Touaibia M. Synthesis and 5-lipoxygenase inhibitory activity of new cinnamoyl and caffeoyl clusters. Bioorg Med Chem Lett 2009; 19:1118–21.
- Loke WM, Proudfoot JM, Stewart S et al. Metabolic transformation has a profound effect on anti-inflammatory activity of flavonoids such as quercetin: lack of association between antioxidant and lipoxygenase inhibitory activity. Biochem Pharmacol 2008; 75:1045–53.
- De la Puerta R, Gutierrez VR & Hoult JRS. Inhibition of leukocyte 5-lipoxygenase by phenolics from virgin olive oil. Biochem Pharmacol 1999; 57:445–9.